

# RASTRUM

## *Protocol*

In Situ Immunofluorescence Analysis



### Introduction

This protocol outlines a method to analyse protein expression via immunofluorescence in 3D cell models created with RASTRUM. This protocol takes three (3) days and is designed for 3D cell models printed into a 96 well plate.

### Equipment and reagents required but not provided

- 4% Paraformaldehyde (PFA)
- Blocking solution
- Phosphate-buffered saline (PBS)
- PBST (0.1% v/v Tween-20 in PBS)
- 0.1% Triton-X-100 in PBS
- Primary (1°) and secondary (2°) antibodies
- Hoechst 33342 (Thermo, cat #H3570)
- Fluorescence microscope

### Protocol

#### Day 1

1. Remove and discard culture medium and add 150  $\mu$ L PBS to each well.
2. Remove PBS and discard and add 100  $\mu$ L 4% PFA to each well
3. Incubate plate at room temperature, 20 minutes.
4. Remove and discard 4% PFA and wash each well with 150  $\mu$ L PBS, three times.
5. Remove and discard PBS and add 100  $\mu$ L 0.1% Triton-X-100 to each well.
6. Incubate plate at room temperature, 30 minutes.
7. Remove and discard Triton-X-100 and wash each well with 150  $\mu$ L PBS, 10 minutes, three times.

**Note:** All waste from Steps 5-8 should be disposed of according to institution-specific laboratory waste disposal guidelines.

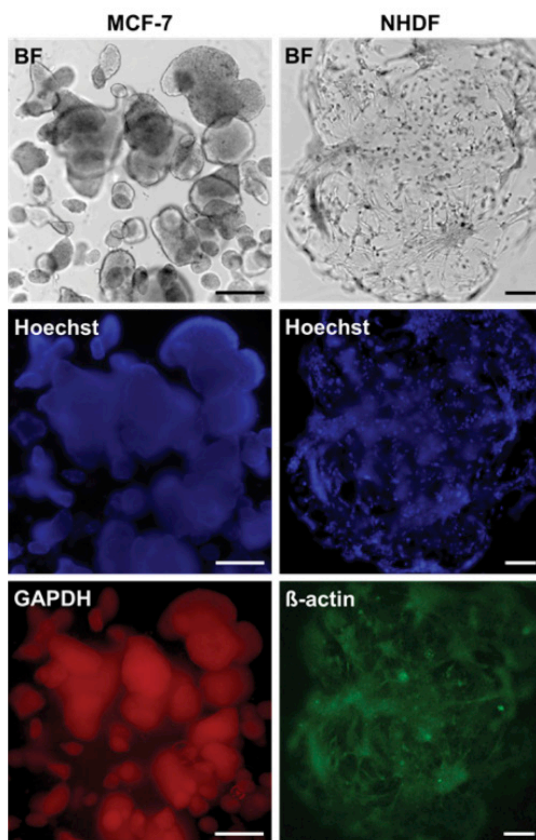
8. Remove PBS and add 100  $\mu$ L blocking solution to each well to block non-specific antibody binding.
9. Incubate plate at room temperature, 30 minutes.
10. Prepare 1° antibody working solution in blocking solution.
11. Remove and discard blocking solution and add 100  $\mu$ L primary antibody working solution to each well.
12. Cover plate with aluminium foil and incubate at 4°C, overnight.

### Day 2

13. Remove and discard 1° antibody and wash each well with 150  $\mu$ L PBST, 10 minutes, three times.
14. Prepare 2° antibody working solution in blocking solution.
15. Remove and discard PBST and add 100  $\mu$ L 2° antibody working solution.
16. Cover plate with aluminium foil and incubate at 4°C, overnight.

### Day 3

17. Remove and discard 2° antibody and wash each well with 150  $\mu$ L PBST, 10 minutes, three times.
18. Prepare 5  $\mu$ g / mL Hoechst 33342 working solution in PBS.
19. Remove and discard PBST and add 100  $\mu$ L Hoechst 33342 working solution.
20. Incubate plate at room temperature, 10 minutes.
21. Remove and discard Hoechst 33342 working solution and add 150  $\mu$ L PBS to each well.
22. Remove and discard PBS wash, and add 150  $\mu$ L PBS to each well.
23. Proceed to imaging with a fluorescence microscope.



**Figure 1:** Representative brightfield (BF), Hoechst and GAPDH/ $\beta$ -actin immunofluorescent images of MCF-7 breast cancer and Neonatal human dermal fibroblast (NHDF) cells encapsulated in RASTRUM hydrogel matrices 7 days post-printing. Scale bars = 200  $\mu$ m



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